Synthesis and evaluation of thiol probes using the 6,7-dimethoxyquinoxalin-2(1*H*)-one fluorophore Shohreh Ghorbanian, Lina K. Mehta and John Parrick*

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3-Substituted quinoxalin-2(1*H*)-ones, **7** and **10**, were prepared as candidate fluorescent probes for thiols and then reacted with representative thiols to give sulfides, **11–17**, whose fluorescence was studied.

The detection and estimation of thiols is an important procedure in environmental and biochemical investigations.¹ The usual reactive centres in fluorescent probes for thiols are maleimide or iodoacetate.^{2–6} We have linked the reactive centre to the fluorophore through a chain conjugated with the quinoxalinone fluorophore so that the reactive centre is bonded to an aromatic nucleus. The molecular architecture was designed to give a degree of flexibility in the chain, an increased fluorescence emission maximum compared with that from a saturated chain, and to provide ease of formation of the maleimide and iodoacetate reactive centres.

Condensation of 6,7-dimethoxy-1,3-dimethylquinoxalin-2(1H)-one, 1,¹³ with 4-acetamidobenzaldehyde and 4nitrobenzaldehyde gave 2 and 4, respectively (Scheme 1). Attempts to reduce the nitro group of 4 without reducing the olefinic function were unsuccessful and only 5 was obtained. The required amine 3 was prepared by hydrolysis of 2. The reaction of 3 with maleimide in chloroform²⁰ gave the maleimic acid 6, which was cyclised to the thiol probe, 7 (Scheme 2). The amine 3 was converted into the haloacetamides 8 and 9 by reaction of haloacetyl halides and 9 was used as a starting material for the preparation of the thiol probe 10. Reaction of butanethiol, thiophenol, 2-hydroxyethanethiol and *N*-acetylcysteine with 7 gave the sulfides 11–14, respectively, and reaction of the first three with 10 gave 15–17.



Fluorescence studies were carried out on 1, 3, 7, 10 and 11–17. The conjugated chain as the substituent on the quinoxalinone nucleus in 3 produced the expected bathochromic shift in the fluorescence emission maximum (λ_{em} for 1 in methanol and dichloromethane: 431 and 427 nm, respectively, but the corresponding values for 3 were 493 and 512 nm). The sulfides 15–17 in methanol had λ_{em} 492–493 and fluorescence quantum yield (ϕ_f) 0.24–0.29 which are little different from the values of λ_{em} 491 and ϕ_f 0.25 obtained for

NRR H₂CO H₂C H₂CC H₂CO Ċн, Ċн, 3 COCH=CHCO, H, R1 = H RR1 = COCH=CHCO NHCOCH.X H₂CO H.CC Ċн 9 X = C110 X = 1





the probe 10. In dichloromethane solution, there was little change in λ_{em} but an increase in ϕ_f to 0.50–0.54 for 15–17 compared with ϕ_f 0.25 for 10.

Methanolic solutions of **7** showed little fluorescence (λ_{em} 495 nm, $\phi_f 0.02$), presumably due to intersystem crossing between the excited π^* state of the fluorophore and the $n \rightarrow \pi^*$ state of the conjugated system of the maleimide.^{7,22,23} Potentially usefully, the thiol adducts **11** to **14** showed a significant increase in both the emission maximum and the quantum yield (λ_{em} 499–504 nm, $\phi_f 0.4$ –0.54).

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Techniques used: ¹H NMR spectroscopy, IR and fluorescence spectroscopy, mass spectrometry, elemental analysis.

References: 23.

Schemes: 2.

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References cited in this synopsis

- R. Cecil and J. R. McPhee, in *Advances in Protein Chemistry*, eds. C. B. Anfinsen Jr., M. L. Anson, K. Bailey and J. T. Edsall, Academic Press, New York, 1959, **14**, 255; P. D. Boyer, in *The Enzymes*, eds. P. D. Boyer, H. A. Lardy and K. Myrbäck, Academic Press, New York, 1959, 2nd ed., vol.1, p. 511.
- 2 M. Machida, M. F. Machida, T. Sekine and Y. Kanaoka, *Chem. Pharm. Bull.*, 1977, **25**, 1678; M. Machida, N. Ushijima, T. Takahashi and Y. Kanaoka, *Chem. Pharm. Bull.*, 1977, **25**, 1289.

- 3 O. S. Wolfbeis and H. Marhold, Monatsh., 1983, 114, 599.
- 4 T. Ueno, S. Hikita, D. Muno, E. Sato, Y. Kanaoka and T. Sekine, *Anal. Biochem.*, 1984, 140, 63.
- 5 C. S. Chaurasia and J. M. Kauffman, *J. Heterocycl. Chem.*, 1990, **27**, 727.
- 6 Y. R. Yang and M. E. Langmuir, J. Heterocycl. Chem., 1991, 28, 1177.
- 7 J. E. T. Corrie, J. Chem. Soc., Perkin Trans. 1, 1994, 2975.
- 13 A. R. Ahmad, L. K. Mehta and J. Parrick, *Tetrahedron*, 1995, 51, 12899.
- 20 M. Z. Barakat, S. K. Shwab and M. M. El-Sader, J. Chem. Soc., 1957, 4133.
- 22 C. W. Wu, L. R. Yarbrough and F. Y. H. Wu, *Biochemistry*, 1976, 15, 2863.
- 23 Y. Kanaoka, Angew. Chem., Int. Ed. Engl., 1977, 16, 137.